## Perspective

# Ribozyme Pharmacokinetic Screening for Predicting Pharmacodynamics Dosing Regimens

### Abstract

There have been only four studies published to date that have addressed locally administered synthetic ribozyme pharmacodynamics in animals. A 2'-O-allyl modified ribozyme targeting amelogenin mRNA inhibited normal enamel formation after submandibular injection in neonatal mice. When injected intraarticularly in the knee, a stabilized ribozyme targeting stromelys in message reduced IL-1-stimulated stromelys in mRNA in rabbit synovium . An anti-protein kinase Ca mRNA targeting ribozyme, comprised totally of 2'-amino-modified ribonucleotide analogs, reduced subcutaneously implanted glioma tumor growth in rats. Finally, an in vitro transcribed ribozyme targeting TNF-a mRNA complexes with cationic lipids injected within the peritoneum decreased lipopolysaccharide-induced TNF-a expression in peritoneal cells obtained by lavage . Although this ribozyme was not chemically modified, use of a cationic lipid formulation likely enhanced the circulation time compared with unformulated ribozyme. Unfortunately, no studies have yet been published which explore the treatment of disseminated disease with systemically delivered synthetic ribozymes.

Keywords: amelogenin • ribozyme •TNF-α mRNA • metastasis • angiozyme

# Introduction

There are two major VEGF receptor subtypes exist in man (KDR and Flt-1) and, though each receptors seem

to be attached growth, their precise roles during this process are poorly understood. As a result of the potential

role of the Flt-1 receptor in regulation tube-shaped structure assembly and its potential involvementin growth we've developed a changed hammerhead ribozyme (ANGIOZYME<sup>™</sup>) that targets Flt-1 RNA for malignant tumor applications The recent advances in ribozyme nuclease stabilization have enabled artificial ribozymes to be considered as potential therapeutic agents [1].

Although several studies have examined the material medica of stabilized oligonucleotides in many species, very little is thought about the pharmacokinetic profile of artificial ribozymes in animal systems. In general, stabilized oligonucleotides are apace distributed following endogenous administration (t1/2a on the order of minutes) with a species and dose dependent ANGIOZYMETM was synthesized in step with the tactic of Wincott et al. ANGIOZYMETM is a 35-mer containing many ester modifications previously shown to reinforce ribozyme stability. For internal labeling with [33P], ANGIOZYMETM was synthesized in two halves [2].

The 3' half-ribozyme was 5' endlabeled using T4 polynucleotide enzyme and  $[\gamma 33P]$  ATP. The two half-ribozymes were then ligated along mistreatment T4 RNA ligase. Full

#### Bekir Karlik\*

University of Qafqaz, Faculty of Engineering, Department of Computer Engineering, Neriman Nerimanov pr.103, Baku, Azerbaijan

\*Author for correspondence: bekirkarlik@beykent.edu.tr

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lung and growth was collected and forthwith frozen on dry ice. Plasma and tissue were hold on at  $-70^{\circ}$ C until analyzed for ANGIOZYMETM concentrations [3].

#### Description

On the top of studies demonstrate however pharmacokinetic evaluation will be wont to facilitate optimize the choice of efficacious dosing regimens. In fact, the combination of pharmacokinetic and pharmacodynamics data is particularly necessary once evaluating novel compounds such as ribozymes. The fast in vivo elimination of

Angiozyme couldn't are foreseen supported its substantial in vitro stability in human humor [4].

#### **Conflict of Interests**

None

#### Acknowledgement

None

#### References

- Agrawal S, Jiang Z, Zhao Q *et al.* Mixed-backbone oligonucleotides as second generation antisense oligonucleotides: In vitro and in vivo studies. *Proc. Natl. Acad. Sci. USA.* 94, 2620-2625 (1997).
- Bijesterbosch MK, Manoharan M, Rump ET et al. In vivo fate of phosphorothioate antisense oligodeoxynucleotides: predominant uptake by scavenger receptors on endothelial liver cells. Nucl. Acids Res. 25, 3290-3296 (1997).
- Crooke ST, Graham MJ, Zuckerman JE *et al.* Pharmacokinetic properties of several novel oligonucleotide analogs in mice. *J Pharmacol. Exp. Ther.* 277, 923-937 (1996).
- DeLong RK, Nolting A, Fisher M et al. Comparative pharmacokinetics, tissue distribution, and tumor accumulation of phosphorothioate, phosphorodithioate, and methylphosphonate oligonucleotides in nude mice. *Antisense Nucleic Acid Drug Dev.* 7, 71-77 (1997).